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# Seed preservatives properties of *Secamone afzelii* (Schult) K. Schum extracts on wheat grains damage and germination capability

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## Abstract

**Background:** Botanical pesticides may be a possible solution to the post-harvest loss of stored grains faced by farmers and food processors. In this study, seed preservatives properties of *Secamone afzelii* leaves extracts at 50, 100, 150, 200  $\mu$ l concentrations were investigated against lesser grain borer *Rhyzopertha dominica* (Fabricus) infestation on wheat grains weight loss and seed germination capability arranged in a Completely Randomized Design (CRD) and replicated thrice under ambient laboratory conditions.

**Results:** The outcomes of the study showed that *S. afzelii* leaves extracts exhibited significant effect ( $p < 0.05$ ) in suppressing *R. dominica* adult emergence and weight loss of wheat grains than untreated. The increase in the concentrations led to significant decrease in adult emergence and weight loss with different extracts. *Secamone afzelii* ethyl acetate and hexane extracts significantly subdued adult emergence in contact toxicity (6.69% and 8.49%) and fumigant toxicity (8.85 and 8.87); a similar trend was observed in weight loss in both contact (2.33% and 5.13%) and fumigant (1.50% and 1.03%) toxicities at 200  $\mu$ l, respectively. None of the extracts hinder the germination of treated grains, while germination percentage ranged between 88.69% and 93.40%.

**Conclusion:** The results clearly indicated that *S. afzelii* extracts, in general, ethyl acetate and hexane extracts of *S. afzelii* in particular were the best seed preservative for stored wheat. Thus *S. afzelii* leaves extracts could be utilized as grain protectant substitute for conventional synthetic insecticides and fumigants in the control of insect infestation and conservation of stored grain quality and germination capability. Further study is required to identify the bioactive chemicals responsible for the *S. afzelii* extracts insecticidal activity and stored grains quality preservation.

**Keywords:** Germination capability, Germination rate, Post-harvest loss, Seed preservatives, Wheat grains

## Background

A number of insect pests induced deleterious effects on cereal grains/seeds post-harvest management. Among the key insect pests of stored cereal grains are the coleopteran beetles, namely: Granary weevil, *Sitophilus granarius* (Linnaeus), maize weevil, *S. zeamais* (Motschulsky), rice weevil *S. oryzae* (Linnaeus), red flour beetle, *Tribolium castaneum* (Herbst), Angoumois grain moth, *Sitotroga cerealella* (Olivier) and Lesser grain borer,

*Rhyzopertha dominica* (Fabricus), and cause substantial grains perforation or damage, weight loss, alteration of nutritional quality, decreased germination potential and leading to colossal economic losses due to reduction in the marketable worth of the seed (Adesina et al. 2019a, b, 2020; Adesina and Mobolade-Adesina 2020).

To protect stored cereal grains against storage insect infestation in storage facilities such as granaries, warehouse/storeroom, rumbus or silo, since it is practically impossible to store food grains/seeds without insect attack, countless number of synthetic insecticides and fumigants remains the conceivable key to the prevention

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of post-harvest loss of wheat and other stored food grains faced by farmers, food grains merchants and processors.

In spite of the effectiveness of synthetic insecticides and fumigants, its uses have faced serious challenges such as increased insecticide residues on stored grains, which pose health threats to human, development of resistant strains and pest resurgence and ecological disorder (Tesfu and Emanu 2013; Khaliq et al. 2014).

Therefore, there is a concerted effort to seek and developed substitute to synthetic insecticides and fumigants to keep stored seed/food grains free of insect pest infestations and for long-term storability and quality parameter maintenance. The use of plant products as botanical pesticides is projected as the most sustainable and ecologically safe tactic to counterbalance the constantly cumulative threat posed by conventional insecticides.

It has been reported that several botanical extracts have the ability of causing negative impact on insect pests (Kestenholz et al. 2007). These extracts have a lot of advantages like readily available and affordable, relatively inoffensive to the environment and to man and offer no residual effects, easy degradable and selective to the targeted pests (Khan et al 2010; Ntonifor et al 2010). In this background, several plant products have been evaluated and considered encouraging in the prevention of stored grain damage and sustenance of seed germination quality.

From available literature, no scientific report of *S. afzelii* products as insecticide against stored wheat insect pests was documented. However, the insecticidal activity of the plant methanol and hexane extracts was effective against *Callosobruchus maculatus* (Adesina and Ofuya 2015). Therefore, this study forms part of the search for bioinsecticides for the management of destructive pests of stored wheat grain products against grain damage and germination capabilities with extracts of *S. afzelii* as the main focus.

## Methods

### Study location and conditions

The experiment was laid out in a Completely Randomized Design (CRD) and replicated thrice in the Pest Management Laboratory, Department of Crop, Pest Management Technology, Rufus Giwa Polytechnic, Owo, Ondo State, Nigeria, under ambient laboratory conditions of  $32 \pm 2$  °C temperature,  $75 \pm 5\%$  relative humidity and 12-h dark/light photoperiods.

### Rearing insect culture

Adults *R. dominica* were obtained from infested wheat grains bought from Ulede market, Owo, Ondo State, Nigeria. Insects were subsequently mass nurtured on whole clean, intact and disinfested wheat in 5-L plastic jars covered with muslin cloth and tightly secured

with rubber band. This was done by weighing 4 kg of wheat grains into clean plastic jars and was infested with about 150 mixed sex adults of *R. dominica* and maintained in the laboratory for mass culture under a temperature of  $32 \pm 2$  °C,  $75 \pm 5\%$  relative humidity and 12-L:12D photoregime. This culture was maintained and used as source of *R. dominica* for all bioassays, and insects of mixed sex required for these trials were 2–4 days old.

### Preparation of *S. afzelii* leaves extracts

Mature vines of *S. afzelii* were collected from Laoso Camp in Ondo, Nigeria. The leaves were identified at the Department of Forestry and Wood Technology, Rufus Giwa Polytechnic, Owo, Ondo State, Nigeria. The leaves were detached and air-dried in a cool dry place for three weeks. The dried leaves were then pulverized into a powder using hammer mill. Thereafter, about 300 g of *S. afzelii* leaves powder was extracted by separately soaking in an extraction bottle containing 500 ml of hexane, ethyl acetate, acetone and methanol respectively for 48 h, occasionally stirred with a glass rod in order to ensure uniformity in extraction. The extracts were strained through a double-layer Whatman's No. 1 filter paper followed by evaporation until the solvent was dried. The crude extracts were stored in vial bottles and stored in the refrigerator for subsequent insecticidal activities by contact and fumigant toxicity (Ileke et al. 2016; Adesina and Mobolade-Adesina 2020; Ileke 2021).

### Contact and fumigant toxicity insect bioassay

About 20 g of the disinfested wheat grains was weighed into each Petri dishes and desiccators for contact and fumigant bioassay, respectively. For contact toxicity, wheat grains were treated with 50, 100 150 and 200  $\mu$ l in triplicates of hexane, ethyl acetate, acetone and methanol extracts of *S. afzelii* in 250-ml plastic dishes using a micropipette. To ensure even mixing of the extracts with the grains, treated grains were thoroughly mixed using a glass rod and then the grains were left to air-dried for about 15 min. After drying of the grains, ten unsexed 2–4-day-old adult *R. dominica* were introduced into each Petri dish. There was also a control treatment containing untreated seeds.

In fumigant toxicity, 10 unsexed 2–4-day-old adult *R. dominica* was exposed to 50, 100, 150 and 200  $\mu$ l of hexane, ethyl acetate, acetone and methanol *S. afzelii* extracts in 0.85-l-L desiccators containing 20 g of wheat grains that served as the fumigation chambers with a Whatman No. 1 filter circle (9 cm size) placed to serve as an evaporating surface for injecting extracts with a micropipette (Adesina et al. 2018). From both experiments, the adult mortality count was recorded for 5 days after infestation and all insects both dead and live were

removed. At 45 days after treatment, number of emerging adult insects in each contact and fumigant toxicity and control were recorded.

The weight loss of wheat grains in the treated and control groups was calculated after 45 days of infestation on a fresh weight basis according to Parkin (1956) formula:  $WI - W/WI \times 100$ , where  $WI$  is the weight of the grains before the experiment and  $W$  is the weight of the grains at the end of the experiment.

### Seed germination test

The effect of *S. afzelii* extracts on germination of treated wheat samples was determined following the method of Adesina et al (2019a). Damaged and undamaged grains carefully separated, 20 g of wheat grains in each of the fumigated and contact toxicities groups were randomly selected from the undamaged ones and placed on moistened tissue paper in sterilized Petri plates as the growth medium and allowed to germinate at room temperature. The germination percentage in fumigated and contact toxicities groups was determined after 24 h. Three replicates were conducted for each sample group. The number of germinated seeds in each treated and control was recorded for all the replicates, and the percent germination was determined (Demissie et al. 2008; Manmathan and Lapitan 2013).

### Statistical analysis

Data obtained from the study were recorded as mean value  $\pm$  SE and subjected to analysis of variance (ANOVA) after the proper transformation, and there were a significant treatment means, Tukey's Student test (HSD) was used to identify differences in treatment means at 5% probability level of significance. All collected data were statistically analyzed using SPSS version 11.5 software.

## Results

### Effect of *Secamone afzelii* extracts on percentage adult emergence

The results presented in Table 1 show that *S. afzelii* extracts were effective in suppressing adult emergence and reducing the population increase of *R. dominica* compared to the untreated control and the level of adult emergence from the treated grains varied with the extracts and concentrations applied. In wheat grains treated with *S. afzelii* extracts at its highest concentrations (200  $\mu$ l/20 g wheat), a total range of 10.26–6.69 adult *R. dominica* insect emergence was recorded, while *S. afzelii* ethyl acetate extracts at its highest concentration recorded the lowest (6.69) adult *R. dominica* emergence. The lowest suppression of *R. dominica* population in grains treated with the extracts was observed at its

**Table 1** Effect of *Secamone afzelii* extracts on percentage adult emergence

Contact toxicity	Concentration ( $\mu$ l)			
	50	100	150	200
Treatments	50	100	150	200
Control	28.93 $\pm$ 1.52a	22.14 $\pm$ 1.50a	26.35 $\pm$ 0.44a	27.06 $\pm$ 0.23a
Hexane	26.23 $\pm$ 2.12a	19.66 $\pm$ 2.04b	13.16 $\pm$ 1.37c	8.49 $\pm$ 1.55b
Ethyl acetate	23.63 $\pm$ 1.38b	20.83 $\pm$ 1.73b	15.28 $\pm$ 2.66b	6.69 $\pm$ 1.03c
Methanol	20.23 $\pm$ 1.78c	15.81 $\pm$ 1.62c	11.81 $\pm$ 1.84d	9.36 $\pm$ 2.31b
Acetone	24.33 $\pm$ 2.91b	16.42 $\pm$ 1.05c	13.81 $\pm$ 2.96c	10.26 $\pm$ 1.88b
<i>Fumigant toxicity</i>				
Control	31.49 $\pm$ 2.88a	38.09 $\pm$ 1.46a	34.40 $\pm$ 1.81a	30.87 $\pm$ 2.55a
Hexane	20.38 $\pm$ 2.61c	15.34 $\pm$ 1.98c	14.70 $\pm$ 2.06c	8.85 $\pm$ 1.50c
Ethyl acetate	28.67 $\pm$ 0.03b	25.00 $\pm$ 3.80b	21.75 $\pm$ 0.30b	8.87 $\pm$ 1.30c
Methanol	27.09 $\pm$ 2.40b	23.69 $\pm$ 4.60bc	17.27 $\pm$ 1.86c	10.74 $\pm$ 3.25b
Acetone	28.86 $\pm$ 1.73b	20.84 $\pm$ 0.81bc	18.55 $\pm$ 1.32c	11.68 $\pm$ 1.38b

Means within a column followed by different letters are significantly different ( $P > 0.05\%$ ), Tukey's Student test (HSD)

**Table 2** Effect of *Secamone afzelii* extracts on percentage weight loss

Contact toxicity	Concentration ( $\mu$ l)			
	50	100	150	200
Treatments	50	100	150	200
Control	28.93 $\pm$ 3.26a	22.14 $\pm$ 2.11a	26.35 $\pm$ 1.94a	27.06 $\pm$ 2.85a
Hexane	13.05 $\pm$ 2.09b	10.94 $\pm$ 0.44b	8.13 $\pm$ 1.16c	5.13 $\pm$ 1.92c
Ethyl acetate	7.71 $\pm$ 2.51c	5.74 $\pm$ 0.38c	3.02 $\pm$ 1.15d	2.33 $\pm$ 1.86d
Methanol	16.32 $\pm$ 2.33b	13.94 $\pm$ 1.88b	14.18 $\pm$ 2.77b	13.94 $\pm$ 0.28b
Acetone	13.18 $\pm$ 2.84b	11.09 $\pm$ 0.54b	9.81 $\pm$ 0.33c	6.55 $\pm$ 1.13c
<i>Fumigant toxicity</i>				
Control	25.40 $\pm$ 0.0a	23.19 $\pm$ 0.55a	22.95 $\pm$ 1.4a	23.42 $\pm$ 1.3a
Hexane	10.71 $\pm$ 0.01b	9.99 $\pm$ 1.20b	7.71 $\pm$ 0.0b	1.03 $\pm$ 0.4c
Ethyl acetate	9.15 $\pm$ 0.03b	7.87 $\pm$ 3.80b	6.09 $\pm$ 0.30b	1.50 $\pm$ 4.70b
Methanol	8.98 $\pm$ 1.40b	8.12 $\pm$ 1.80b	7.47 $\pm$ 1.10b	6.20 $\pm$ 2.20b
Acetone	9.99 $\pm$ 2.50b	3.84 $\pm$ 1.80c	2.11 $\pm$ 1.80c	4.87 $\pm$ 0.66c

Means within a column followed by different letters are significantly different ( $P > 0.05\%$ ), Tukey's Student's test (HSD)

highest dosage in acetone extract (10.26). However, none of the extracts were able to achieve complete suppression of adult emergence.

### Effect of *Secamone afzelii* extracts on percentage weight loss

The percentage weight loss has a very highly significant variation following extracts and concentrations (Table 2). The highest weight loss (28.93 and 25.40%) was recorded from untreated grains compared to treated grains, which have better protection that recorded a significant low

weight loss. The percentage weight loss decreased with the increasing of the concentration level. The percentage weight loss among different concentrations of the *S. afzelii* leaf extracts was lowest in contact toxicity of wheat treated with 200 µl ethyl acetate extract (2.33%) followed by a hexane extract (5.13%), while in fumigant toxicity, the lowest weight loss was recorded in hexane extract (1.03%) and closely followed by ethyl acetate extract (1.50%).

**Effect of *Secamone afzelii* extracts on percentage seed germination**

The results in Table 3 show a significant encouraging effect of *S. afzelii* extracts on the germination percentage of wheat grain after treatments and storage. A non-significant difference ( $p < 0.05$ ) was observed between wheat grains in controls and the different concentrations of the extracts. The wheat grains treated with the lowest concentration of the extracts (5 µl/20 kg wheat) and untreated (control) recorded more or less same germination rate. From this result, it is evident that seeds preserved with the different concentrations of the extracts did not lose their viability.

**Discussion**

Grain protectants play vital role in extending the storability of seeds to longer duration without appreciable loss in vigor and viability (Adesina et al. 2019a). The findings obtained in this study showed that *S. afzelii* extracts are effective as stored grains protectant against storage

insects in reducing grain damage and ultimately lower weight loss.

The present result complemented the outcomes of various study by researchers who assessed diverse plant extracts on most destructive stored product insect pests and establish that insects were highly vulnerable to contact and fumigant actions of botanical extracts and thus act as a good substitute to high continuing synthetic insecticides and fumigants for dealing with the insect pests in stored agricultural produce (Ukih et al 2010; Waseem et al 2019).

All the extracts treatments induced significant reduction in *R. dominica* adult emergence compared to the untreated control, although the extracts adult suppression potentiality varies within the treatments. *Secamone afzelii* ethyl acetate and hexane extracts were superior in reducing the F1 progeny emergence, and *S. afzelii* acetone extract was less effective compared to other extracts. The reduction in adult emergence observed in the treated grains might be as a result of the low survival rate of parent adult insect exposed to contact and fumigant toxicity, anti-oviposition, ovicidal and larvicidal properties of the tested extracts. This aligned with the findings of Taponjou et al. (2002); Adesina et al. (2011); Adesina and Ofuya (2015); Adesina et al. (2015a, b) who opined that the oviposition inhibition property of plant products on adult storage insects makes them laid fewer eggs and where eggs are hatched the plant products killed the larvae hatching from eggs laid on grains. Udo (2005) opined that there is a relationship between first filla adult insect emergence and parental death as well as the likely presence of oviposition restrictive. Decline in offspring emergence from grains treated with *S. afzelii* extracts might be as a result of early mortality of adult insect and fractional or complete hindrance of embryonic growth. The significant reduction of adult emergence obtained from the study could also be due to the reaction of the different bioactive compounds present in these extracts.

The findings obtained in this study agreed with the earlier reports that plant extracts could adequately protect stored wheat from attack of its major insect pest of *R. dominica* owing to its F1 progeny production significant suppression. The ability of a botanical to act as biopesticides is not only accessed by its ability to evoked adult mortality but by its capability to hinder offspring development in treated grains (Khoshnoud et al. 2008).

The reduction in seed damage as manifested in the low weight loss recorded could be due to the likely biochemical constituents present in *S. afzelii* extracts which invariably caused adult mortality, reduction of adult emergence, hindered insect developmental stages, insect feeding on stored grains and seed damage. The present results substantiate the findings of Govindan and Nelson

**Table 3 Effect of *Secamone afzelii* extracts on percentage seed germination**

Contact toxicity	Concentration (µl)			
	50	100	150	200
Treatments	50	100	150	200
Control	88.69 ± 1.52a	90.55 ± 1.50a	91.69 ± 0.44a	92.40 ± 0.23a
Hexane	91.95 ± 1.20a	89.11 ± 1.17a	90.69 ± 0.8a	93.40 ± 0.38a
Ethyl acetate	90.24 ± 0.83a	91.40 ± 1.83a	90.84 ± 1.06a	90.25 ± 2.06a
Methanol	89.39 ± 0.52a	93.40 ± 1.62a	90.69 ± 0.48a	91.95 ± 1.55a
Acetone	92.24 ± 1.48a	90.26 ± 1.50a	90.40 ± 1.59a	91.66 ± 1.88a
Fumigant toxicity				
Control	90.00 ± 0.0a	91.39 ± 0.55a	90.00 ± 1.4a	90.00 ± 1.3a
Hexane	90.30 ± 0.33a	90.88 ± 1.22a	91.39 ± 0.55a	92.00 ± 0.74a
Ethyl acetate	90.00 ± 0.82a	91.69 ± 0.85a	89.69 ± 0.94a	90.80 ± 0.22a
Methanol	92.69 ± 0.76a	91.30 ± 1.88a	91.39 ± 1.05a	90.00 ± 0.38
Acetone	90.30 ± 0.33a	90.69 ± 0.57a	90.69 ± 0.57a	90.00 ± 0.38

Means within a column followed by different letters are significantly different ( $P > 0.05\%$ ), Tukey's Student test (HSD)



(2008); Adesina and Ofuya (2015) and Chandrakala et al. (2013) who reported the reduction in weight loss of stored grains when treated with different plant products and that plant products may be a prospective source of biopesticide for use in pest control approaches in preservation of stored grains against insect attack. Adesina et al. (2019b) stated that substantial infestation and associated grains damage and weight loss noted from the untreated grains evidently showed that vulnerable grains recorded high F1 progeny emergence and predisposed the grains to increasing percentage seed damage and ultimately weight loss due to the unhindered feeding, developmental and metabolic activities of the insects within the stored grains. The outcome of this study tallies with Adesina et al (2011); Adesina et al (2012); Adesina and Mobolade-Adesina (2016) who stated that postharvest weight loss of stored grains correlated with adult emergence inhibition rate.

Several botanical pesticides have been reported to affect the seed germination and seedling growth from treated seeds (Bell 1994; De Groot 1996; Seignobos 2002). The seeds used for germination test in this study were not damaged nor showed evidence of adult exit holes, though they could contain the hidden immature stages of weevil that could destroy the parts of seed embryo. However, in the present investigation, *S. afzelii* extracts did not harmfully affect the sprouting of the treated wheat as the germination of treated wheat did not exhibit any significant difference compared to untreated wheat which shows that *S. afzelii* extracts do not exhibit any phytotoxic effects on the seed. This property of the extracts backs its safety for use in maintaining the quality of stored grain for seed purpose and controlling the stored beetle infestation. This study established previous reports that plant products could effectively protect stored grains against storage insects (Paul et al. 2009) and conserved seed viability without having any negative consequence on germination (Mishra and Dubey 1994; Keita et al. 2001; Kishore and Dubey 2002; Goudoungou et al. 2015; Danjumma et al. 2018; Tagne et al. 2018; Adesina et al 2019a, c).

## Conclusion

In conclusion, the present study indicated that leaves extracts of *S. afzelii* demonstrated significant botanical insecticidal action in the protection of stored wheat grains by enhancing varying degrees of adult emergence inhibition, feeding deterrence and reduced grain damaged as well as weight loss. Also, the treatments did not show any noticeable adverse effects on the germination capacity of the wheat seeds. Therefore, leaves extracts of *S. afzelii* can be utilized as a green insecticide substitute to conventional synthetic insecticides and fumigants

in the control of insect infestation and preservation of stored grain quality. The plant is readily and widely available in many agro-ecological zones of Nigeria and used in the treatment of many ailments in folk medicine. Identification of the bioactive chemicals responsible for the *S. afzelii* extracts insecticidal activity and stored grains quality preservation should be a speedy research focus. Also, the toxicity of the plant extracts on albino rats should be evaluated.

## Abbreviations

ANOVA: Analysis of variance; HSD: Honest significant difference; CRD: Completely Randomized Design.

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## Authors' contributions

JMA conceived, designed and carried out the experiment and prepared the draft manuscript, and ATBA contributed to experiment conception and design, carried out statistical analysis and interpretation of data and proof-readed the manuscript. All authors read and approved the final manuscript.

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## Availability of data and materials

Sharing of data is not applicable.

## Declarations

## Ethics approval and consent to participate

Not applicable.

## Consent for publication

Not applicable.

## Competing interests

The authors have no competing interests to report.

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